## IN THE CLAIMS

- Claim 1 (original): A method for examining the activity of ion channels, comprising the following steps:
  - providing a sample comprising ion channels; and
  - determining a value of a measuring parameter as an indicator of the activity of the ion channels, the measuring parameter being a membrane potential, a measure of a membrane potential, an ion concentration, or a measure of an ion concentration;

characterised in that said determining of the value of the measuring parameter is performed at a temperature of  $\leq$  about 10 °C by fluorescence methods, radioactive methods or atomic absorption spectroscopy.

- Claim 2 (original): The method according to claim 1, characterized in that said determining of the value of the measuring parameter is performed at a temperature of ≤ about 5 °C, especially ≤ about 2 °C.
- Claim 3 (currently amended): The method according to claim 1 or 2, characterized in that said determining of the value of the measuring parameter is performed at a temperature of from about 10 °C to -4 °C, especially from about 5 °C to -4 °C, more preferably from about 5 °C to 0 °C, even more preferably from about 2 °C to 0 °C.
- Claim 4 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the sample comprises one or more cells or cell organelles which have ion

- channels, in particular human or animal cells or cell organelles.
- Claim 5 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the sample comprises one or more vesicles which have ion channels.
- Claim 6 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the sample comprises membrane bound ion channels, in particular ion channels embedded into a membrane of cells, cell organelles, vesicles or embedded into an artificial membrane.
- Claim 7 (currently amended): The method according to any of the preceding claims claim 1, characterized in that said measuring parameter is the membrane potential of a cell, cell organelle or vesicle, or a measure of said membrane potential.
- Claim 8 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the measuring parameter is an extracellular, intracellular, extravesicular and/or intravesicular ion concentration or a measure thereof.
- Claim 9 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the value of said measuring parameter is determined before, during and/or after the addition of a test substance which potentially influences the activity of the ion channels.
- Claim 10 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the activity of a transmitter-dependent ion channel is examined.
- Claim 11 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the activity of

- a voltage-sensitive ion channel is examined.
- Claim 12 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the activity of a potassium channel, chloride channel, sodium channel or calcium channel is examined.
- Claim 13 (currently amended): The method according to any of the preceding claims claim 1 characterized in that an optical response of (i) a carbocyanine derivative, in particular a thia-, indo-, or oxa-carbocyanine or an iodide derivative of a carbocyanine, (ii) a rhodamine dye, (iii) an oxonol dye, (iv) merocyanine 540, or (v) a styryl dye serves as a measure of the membrane potential.
- Claim 14 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the fluorescence emission of a voltage-sensitive fluorescent dye, preferably a DiBAC dye, more preferably the dye Dibac4(3), serves as a measure of the membrane potential.
- Claim 15 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the ion concentration of rubidium, especially of non-radioactive rubidium, is determined as an indicator of the activity of the ion channels.
- Claim 16 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the ion concentration, especially the ion concentration of calcium, is measured by means of chelating agents.
- Claim 17 (currently amended): The method according to any of the preceding claims claim 1, characterized in that the values of several measuring parameters are determined.

- Claim 18 (currently amended): The method according to any of the preceding claims claim 1 for use in the research on pharmaceutically active substances, especially in the medium-or high-throughput screening of potentially or established active pharmaceutical substances, in particular the identification of potentially active pharmaceutical substances or the determination of side effects of potentially or established active pharmaceutical substances.
- Claim 19 (currently amended): The method according to any of the preceding claims claim 1 for use in the agricultural research, especially in the research on agrochemicals as e.g. insectizids.
- Claim 20 (currently amended): Use of a voltage-sensitive or ion-sensitive indicator for the conductance of the method according to any of the preceding claims claim 1.
- Claim 21 (original): Use according to claim 20 wherein the ionsensitive indicator is a calcium indicator, in particular a fluo-calcium indicator, a fura indicator, an indo indicator, Calcium Green™, or Oregon Green™.
- Claim 22 (original): Use according to claim 20 wherein the ion-sensitive indicator is a sodium or potassium indicator, preferably a fluorescent sodium or potassium indicator, in particular SBFI, PBFI, Sodium Green Na<sup>+</sup> indicator, CoroNa Green Na<sup>+</sup> indicator, or CoroNa Red Na<sup>+</sup> indicator.
- Claim 23 (original): Use according to claim 20 wherein the voltagesensitive indicator is a carbocyanine derivative, in particular an indo-, thia-, or oxa- carbocyanine or a iodide derivative of a carbocyanine; a rhodamine dye; an oxonol dye; merocyanine 540; or a styryl dye.

- Claim 24 (original): Use according to claim 23 wherein the oxonol dye is a bis-isoxazolone oxonol dye or a bis-barbituric acid oxonol (DiBAC) dye, in particular  $DiBAC_4(3)$ ,  $DiSBAC_2(3)$  or  $DiBAC_4(5)$ .
- Claim 25 (original): Use according to claim 23 wherein the styryl dye is an ANEP (AminoNaphthylEthenylPyridinium) dye, in particular di-4-ANEPPS, di-8-ANEPPS, di-2-ANEPEQ, di-8-ANEPPQ, di-12-ANEPPQ, di-1-ANEPIA, or a dialkylaminophenylpolyenylpyridinium dye (RH dye), in particular RH 414, RH 421, RH 795 or RH 237.
- Claim 26 (currently amended): Use of a chelating agent for the conductance of the method according to any of claims 1 to 19 claim 1.
- Claim 27 (currently amended): Use of rubidium, in particular non-radioactive rubidium, for the conductance of the method according to any of claims 1 to 19 claim 1.
- Claim 28 (currently amended): Use of an atomic absorption spectrometer, a flow cytometer, a fluorescence microcope or fluorescence plate reader for the conductance of the method according to any of claims 1 to 19 claim 1.
- Claim 29 (currently amended): Use according to claim 28 of an atomic absorption spectrometer, a flow cytometer, a fluorescence microcope or fluorescence plate reader for applying a voltage-sensitive or ion-sensitive indicator according to any of claims claim 20 to 25, a chelating agent according to claim 26 and/or rubidium according to claim 27.

Claim 30 (new): Use of an atomic absorption spectrometer, a flow cytometer, a fluorescence microcope or fluorescence plate reader for applying a chelating agent according to claim 26.

Claim 31 (new): Use of an atomic absorption spectrometer, a flow cytometer, a fluorescence microcope or fluorescence plate reader for applying rubidium according to claim 27.